

Length heteroplasmy in the HVI control region

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Abstract. The variability in np16184–16193 HVI region presents two different types of polymorphism: sequence variability and length variability with length heteroplasmy. It is important to define properly the HVI homopolimeric regions for a correct classification of homopolimeric track defining the number of As and Cs and the various population types identified by adjacent bases to the homopolimeric track. © 2005 Elsevier B.V. All rights reserved.

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1. Introduction

MtDNA is generally accepted as a tool for forensic identity testing and evolutionary studies. It is used to analyse biological samples where the quality and/or quantity of nuclear DNA content is low. In order to properly interpret results, one needs to appreciate mtDNA features such as nomenclature, heteroplasmy, recombination and haploid maternal inheritance. Forensic community has adopted a common language to describe the variation observed in human populations naming mtDNA sequences by referring to a standard sequence—Cambridge Reference Sequence.

HVI mtDNA region have one cytosine segment at np16184–16193, which in accordance with CRS possessed a thymine at np16189. However, some mtDNA sequences have a transition T–C at np16189 or show some kind of sequence variability which is characterized by the number and position of Cs and Ts (CxTx) in that region [1,2].

The variability in np16184–16193 HVI region presents two different types of polymorphism: sequence variability considering distinct thymine positions not interfering with results and length variability with length heteroplasmy with no thymine position in this region giving rise to complex patterns in this area. So it is important to define properly the HVI homopolimeric regions [3] for a correct classification of homopolimeric track

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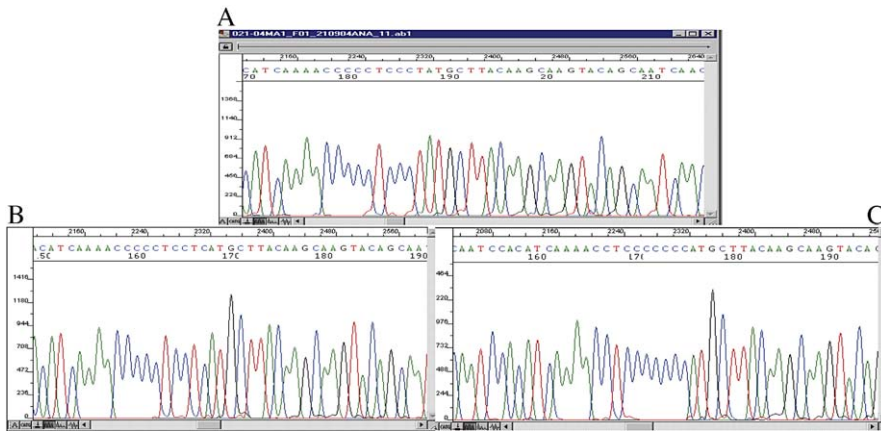


Fig. 1. Sequence variability at HVI np 16183–16193 region characterized by the number and position of timines T (CxTx) at this poli-C mtDNA region: A—C5TC3T; B—C5TC2TC; C—C2TC7.

defining the number of As and Cs and the various population types identified by adjacent bases to the homopolimeric track.

2. Materials and methods

A population study was performed in 98 maternally unrelated individuals from South Portuguese population. DNA was extracted from bloodstains with Chelex 100. HVI mtDNA region was amplified in a 9700 GeneAmp Thermal Cycler (Perkin Elmer) using HVI primers L15997(A1)/H16395(B1) [4]. PCR was carried out in a 25 ul reaction mixture consisting of 2.5 ul GeneAmp 10× Gold Buffer, 1.5 mM MgCl₂, 200 μM dNTPs, 100 μM each primer, 2 U of Taq Gold. The amplification was carried out at 95 °C for 11 min, followed by 36 cycles of 95 °C for 10 s, 60 °C for 30 s and 72 °C for 30 s and a final extension of 72 °C for 10 min. Prior to sequencing, PCR products were purified with Microcon YM100®.

Sequencing fragments were generated using Bigdye® Terminator v1.1 Cycle Sequencing Kit (Applied Biosystems). Sequencing was carried out at 96 °C for 1 min followed by 25 cycles of 96 °C for 10 s, 50 °C for 5 s and 60 °C for 4 min. Removal of excess products was accomplished with Ethanol/EDTA as recommended by the Bigdye®

Table 1
Polymorphism at np16181–16193 with a T–C transition at np16189

CRS	181	182	183	184	185	186	187	188	189	190	191	192	193	Nomenclature	Sample number
	A	A	A	C	C	C	C	C	T	C	C	C	C		
Hc1I									C					A3C10	4
Hc2I									C			.1C		A3C11	5
Hc3I									C			.2C		A3C12	3
Hc4I									C			.3C		A3C13	1
Hc5I			C						C					A2C11	4
Hc6I			C						C			.1C		A2C12	6
Hc7I			C						C			.2C		A2C13	4

