

A comparative study of the sensitivity and specificity of luminol and fluorescein on diluted and aged bloodstains and subsequent STRs typing

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Abstract. Luminol and fluorescein are very important reagents for diluted and aged Bloodstain detection at crime scene. The aim of this study was to carry out a comparative study of the sensitive and selectivity of these two presumptive blood tests using a series of diluted blood (from 1:10 to 1:10.000.000) on a large variety of substrates, as well as, to evaluate the ability to type STRs on treated samples. © 2006 Published by Elsevier B.V.

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1. Introduction

Presumptive reagents, such as luminol and fluorescein, are largely used to detect dilute blood that can not be seen by eye. Although their utility is undoubted, in deciding the optimal reagent to use is desirable to appreciate their utility and limitations, considering false positive reactions, dilution of traces, sensibility on extremely diluted trace and last but not least, limitations for DNA typing, using the most recent STRs markers commercially available.

2. Materials and methods

Blood from a single donor, drawn by venipuncture into an EDTA tube was diluted 1:10, 1:1.000, 1:100.000, 1:10.000.000 with physiological buffer. These four groups of solutions were then called Co1, Co2, Co3 and Co4. One hundred ul aliquots of diluted blood were

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then deposited on nine different substrates (labelled from A to I, Table 1), and allowed to dry and age for 3 weeks. Luminol and fluorescein (in the reduced state) were prepared fresh according to Budowle et al. [1,2] and subsequently applied on the entire surface of the stains. Luminol-treated stains were viewed in complete dark with naked eye, while fluorescein stained materials were viewed using an alternate light source (Crimescope CS16 Spex Yobin-Ivon), set at 450 nm. Bloodstains were collected both by cutting or by swabbing with “OralSwab” (Whatman) wetted with distilled water. DNA extraction was performed with the commercial kit “ChargheSwitch Forensic DNA Purification Kit” (Invitrogen) according to the manufacturer’s protocol. Genotyping was carried out by using AmpF/STR® Identifier™ DNA Amplification Kit from Applied Biosystems and PowerPlex 16.2 kit (Promega) according to the protocol provided by the suppliers. Results were analysed with GeneMapper v.3.2 (ABD). Due to the results obtained by DNA quantification, PCR amplification was limited only to the dilutions set Co1 and Co2.

3. Results and discussion

The detection ability of both reagents is shown in Table 1.

Results of genetic analyses are shown in Tables 2 and 3.

Table 1

+ detectable, – not detectable

| | Luminol | | | | Florescein | | | |
|------------|---------|-----|-----|-----|------------|-----|-----|-----|
| | Co1 | Co2 | Co3 | Co4 | Co1 | Co2 | Co3 | Co4 |
| Wood | + | + | + | – | + | + | – | – |
| Stone | + | + | + | + | + | + | + | + |
| Iron | + | + | – | – | + | + | + | + |
| Plastic | + | + | + | + | + | + | + | + |
| Rubber | + | + | + | – | + | + | – | – |
| Wax-cloth | + | + | + | + | + | + | + | + |
| Baked clay | + | + | + | – | + | + | + | + |
| Linoleum | + | + | + | + | + | + | + | + |
| Cotton | + | + | – | – | + | + | – | – |

Table 2

DNA quantification and profile quality (n.d., not detected, *stochastic effects) of sample from stain Co1 diluted

Dilution 1:10 (Co 1)

| Sample | Substrate | Real-time PCR (ng/μl) | | Quantiblot (ng/μl) | | RFU of the DNA profiles | |
|--------|------------|-----------------------|-------|--------------------|-------|-------------------------|-----------|
| | | LUM | FLU | LUM | FLU | LUM | FLU |
| 1A | Wood | 0.019 | 0.024 | 0.052 | n.d. | 0–400* | 0–300* |
| 1B | Stone | 0.440 | 1.106 | n.d. | 0.247 | 500–5000 | 300–2500 |
| 1C | Iron | 0.339 | 0.472 | n.d. | 0.070 | 200–2000* | 2000–5000 |
| 1D | Plastic | 3.774 | 2.332 | 0.634 | 0.463 | 3000–8000 | 3000–7000 |
| 1E | Rubber | 0.608 | 1.880 | 0.153 | 0.531 | 2000–6000 | 3000–7000 |
| 1F | Wax-cloth | 2.099 | 1.190 | 0.279 | 0.253 | 1000–5000 | 2000–6000 |
| 1G | Baked clay | 2.727 | 1.023 | 0.256 | 0.286 | 2000–6000 | 800–3000 |
| 1H | Linoleum | 1.390 | 0.447 | 0.228 | 0.081 | 2000–6000 | 4000–9000 |
| 1I | Cotton | 1.031 | 0.634 | 0.063 | 0.660 | 2000–7000 | 500–3000 |

Table 3

DNA quantification and profile quality (n.d., not detected, *stochastic effects) of sample from stain Co2 diluted

Dilution 1:1.000 (Co 2)

| Sample | Substrate | Real-time PCR (ng/μl) | | Quantiblot (ng/μl) | | RFU of the DNA profiles | |
|--------|------------|-----------------------|-------|--------------------|-------|-------------------------|-----------|
| | | LUM | FLU | LUM | FLU | LUM | FLU |
| 2A | Wood | 0.013 | 0.003 | n.d. | n.d. | 100–600* | 100–600* |
| 2B | Stone | 0.006 | 0.040 | n.d. | n.d. | 100–300* | 200–800* |
| 2C | Iron | n.d. | n.d. | 0.043 | n.d. | 0–500* | 0–250* |
| 2D | Plastic | 0.179 | 0.087 | 0.048 | n.d. | 200–1500 | 200–2000* |
| 2E | Rubber | 0.126 | 0.088 | n.d. | 0.056 | 400–2000 | 500–2000* |
| 2F | Wax-cloth | 0.107 | 0.198 | n.d. | 0.051 | 100–2000* | 1000–4000 |
| 2G | Baked clay | 0.082 | 0.059 | 0.081 | n.d. | 100–500* | 100–1000* |
| 2H | Linoleum | 0.153 | 0.035 | 0.034 | n.d. | 100–500* | 100–600* |
| 2I | Cotton | 0.050 | 0.056 | 0.064 | 0.036 | 100–300* | 300–1500 |

As expected, luminol and fluorescein have shown different detection capabilities depending upon the substrate. Luminol compared to fluorescein seems to be more sensitive on wood and rubber (Co3 instead of Co2 dilution) while fluorescein has shown more sensitivity on iron and baked-clay (Co4 respectively instead of Co2 and Co3). As it regards stone, iron, plastic and wax-cloth, both reagents have shown the same huge sensitivity, till the last dilution Co4. Cotton, instead, gave different results, as we were able to detect bloodstains just until Co2. Our results confirm that non-absorbent substrates offer a better surface to detect blood, independently of the reagents used as showed by Budowle et al. [1]. Relevant DNA typing has demonstrated that it is possible to have a blood dilution up to 1:1000, corresponding to a DNA quantity of about 50 pg/μl, independently of the reagent used to treat the sample, showing results never seen in previous studies. Even if no statistical tests have been conducted, no significant difference, regarding the DNA extraction as well as the capability to be amplified, can be deducted from the dataset. Independently of the presumptive reagent used, it was not possible to obtain typeable results from extremely diluted bloodstains (Co3 and Co4, data not shown). Stochastic effects, such as drop-out or missing genotype have occurred during the PCR of Co2 diluted samples, due to the very low quantity of DNA rather than being affected by the luminol or the fluorescein treatment. In conclusion, the results of the current study confirm that either luminol or fluorescein are excellent reagents to detect latent and aged bloodstain at crime scene. They also showed that their use does not interfere with the ability to recover typeable DNA. Using the most recent STRs markers, it was possible to obtain full DNA profiles even at very high blood dilutions (i.e. 1:1000), obtaining results never achieved in the past, which encourage their use for criminal investigation.

References

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