

AmpF ℓ STR[®]Y-filer[™]: A new tool for rapid Y-STR forensic haplotyping

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Abstract. Allele frequencies and population data for 17 Y-STR loci, including all the markers in the actually used European “extended haplotype” (DYS19, DYS189I, DYS389II, DYS390, DYS391, DYS392, DYS393, DYS385I/II, DYS438 and DYS439) and also DYS437, DYS448, DYS456, DYS458, DYS635 and Y GATA H4 included in a new commercial kit that has recently been available, the AmpF ℓ STR[®] Y-filer[™] PCR amplification kit (Applied Biosystems), were obtained from a sample of 175 healthy unrelated males and 45 father–son pairs from the North of Portugal. A total of 171 haplotypes were identified, of which 167 were unique and 4 found in two individuals. The haplotype diversity (99.97%) and discrimination capacity (95.43%) were calculated. © 2005 Published by Elsevier B.V.

Keywords: Y-chromosome; Short tandem repeat typing; Multiplex amplification; North of Portugal; Y-filer[™]

1. Introduction

In addition to the standard panel of autosomal loci used in forensic genetics, Y-STR haplotyping gives the ability to sensitive typing of male-specific DNA especially in sexual assault cases or other situations where mixtures of male and female cells are present [1]. Within the last years, a number of Y-STR multiplex assays have been developed [2]. Recently, a new commercial kit has been available, the AmpF ℓ STR[®] Y-filer[™] PCR amplification kit (Applied Biosystems) that permits the simultaneous amplification of 17 Y-STR loci, including all the markers in the actually used European “extended haplotype”,

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Table 1
Allele frequencies at 17 Y-STRs in a North of Portugal population sample (175 individuals)

Allele	DYS19	DYS389I	DYS389II	DYS390	DYS391	DYS392	DYS393	DYS437	DYS438	DYS439	DYS448	DYS456	DYS458	DYS635	GATA H4	Genotypes	DYS385
8									0.006	0.040				0.029		10–14	0.006
9					0.051				0.114	0.337				0.206	0.011	11–11	0.017
10					0.503	0.006			0.291	0.469				0.086	0.017	11–12	0.023
11		0.006			0.423	0.360			0.051	0.143				0.520	0.320	11–13	0.040
12	0.006	0.166			0.023	0.057	0.206		0.509	0.006			0.006	0.120	0.583	11–14	0.303
13	0.120	0.657				0.537	0.651		0.029			0.011	0.000	0.040	0.069	11–15	0.114
14	0.566	0.166				0.040	0.120	0.366				0.097	0.046			12–12	0.029
15	0.229	0.006					0.023	0.514				0.457	0.097			12–13	0.006
16	0.063							0.114				0.314	0.200			12–14	0.034
17	0.017							0.006			0.006	0.097	0.303			12–15	0.034
18											0.086	0.023	0.211			12–17	0.006
19											0.486		0.114			12–20	0.006
20											0.246		0.023			13–14	0.051
21				0.029							0.137					13–15	0.046
22				0.103							0.029					13–16	0.034
23				0.194							0.011					13–17	0.040
24				0.554												13–18	0.006
25				0.091												13–19	0.011
26			0.023	0.017												14–14	0.011
27			0.029	0.006												14–15	0.029
28			0.131													14–16	0.023
29			0.480													14–17	0.011
30			0.246													14–18	0.006
31			0.080													14–19	0.006
32			0.029													15–15	0.017
R										0.006						15–16	0.006
																15–17	0.006
																15–18	0.011
																16–16	0.006
																16–17	0.011
																16–18	0.023
																16–19	0.011
																16–20	0.006
																17–17	0.006

R corresponds to a duplicated allele—10–12.

DYS19, DYS189I, DYS389II, DYS390, DYS391, DYS392, DYS393, DYS385I/II, DYS438, DYS439 and also DYS437, DYS448, DYS456, DYS458, DYS635 and Y GATA H4. Here we report population data for a sample of 175 males and 45 father–son pairs from the North of Portugal.

2. Materials and methods

Blood and/or oral swabs from 175 apparently healthy and unrelated males and 45 father–son pairs from the North of Portugal were collected. DNA was extracted using the Chelex or a standard phenol–chloroform method. The loci DYS19, DYS389I and II, DYS385, DYS390, DYS391, DYS392, DYS393, DYS456, DYS458, DYS437, DYS438, DYS439, DYS448, DYS635 and Y GATA H4 were co-amplified by using the AmpF ℓ STR $^{\text{®}}$ Y-filer $^{\text{™}}$ PCR amplification Kit (Applied Biosystems) following the conditions recommended by the manufacturer (Technical Manual) and carried out using a thermocycler GeneAmp $^{\text{®}}$ PCR System 9700 (Applied Biosystems). The ABI PRISM $^{\text{®}}$ 3100 Genetic Analyzer (Applied Biosystems) was used for genetic typing. Fragment sizes were determined automatically using the Genescan $^{\text{®}}$ Analysis Software v. 3.7 and allele designations were based on comparison with allelic ladders included in the Y-filer $^{\text{™}}$ kit. We also used some known control samples for comparison. Allele frequencies were estimated by gene counting. Gene and haplotype diversities were calculated according to Nei [3] using the Arlequin Software, version 2000 [4] and the discrimination capacity was also determined.

3. Results and discussion

Allele frequencies for the 17 Y-chromosome STRs are shown in Table 1. A total of 171 different haplotypes were identified, 167 of them were unique and 4 were found in two individuals. The observed haplotype diversity was 0.9997 ± 0.0007 . In the system DYS439, we have found a duplicated allele (10–12) that needs to be sequenced. The generated data shows that the haplotype constructed using the present set of 17 Y-STR markers studied with the AmpF ℓ STR $^{\text{®}}$ Y-filer $^{\text{™}}$ kit is highly polymorphic and discriminative in the North of Portugal population. The genotyping with this kit resulted in a greater discrimination capacity when compared with the PowerPlex $^{\text{®}}$ Y System kit (Promega Corporation) (data not shown). This is important when regarding forensic casework analysis since it permits the genotyping of a greater set of markers using less DNA sample. No comparisons were made with other populations due to the lack of information for these particular set of loci.

In 2 of the 45 father–son pairs analysed, we have found inconsistencies of allelic transmission in the loci DYS448 and Y DYS456, which must be confirmed by sequencing.

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